



Mushroom Cultivation Systems: Exploring Antimicrobial and Prebiotic Benefits

Xhensila Llanaj^{1,2}, Gréta Törös^{1,3}, Péter Hajdú^{1,2}, Hassan El-Ramady^{1,4}, Ferenc Peles⁵ and József Prokisch¹

¹*Institute of Animal Science, Biotechnology and Nature Conservation, Faculty of Agricultural and Food Sciences and Environmental Management, University of Debrecen, 138 Böszörményi Street, 4032 Debrecen, Hungary*

²*Doctoral School of Food Science, University of Debrecen, Böszörményi Street 138, 4032 Debrecen, Hungary*

³*Doctoral School of Animal Husbandry, University of Debrecen, Böszörményi Street 138, 4032 Debrecen, Hungary*

⁴*Soil and Water Dept., Faculty of Agriculture, Kafrelsheikh University, 33516 Kafr El-Sheikh, Egypt*

⁵*Institute of Food Science, Faculty of Agricultural and Food Sciences and Environmental Management, University of Debrecen, Böszörményi Street 138, H-4032 Debrecen, Hungary*

MUSHROOM farming is the practice of growing and cultivating various species of mushrooms for food, medicinal, or industrial purposes. It involves creating the ideal environment and conditions for the mushrooms to grow and thrive, such as providing a substrate (a nutrient-rich material on which mushrooms grow), controlling temperature and humidity, and ensuring proper ventilation and lighting. Several innovative applications of mushrooms can be noticed in their farming and cultivation. These applications mainly focus on the medicinal and prebiotic attributes, besides their role in producing food and energy. This review is an attempt to highlight the cultivation of mushrooms and their requirements under different kinds of farming. These farming types may include mushroom-worm system, mushroom-bee farming, smart mushroom farming, forest-mushrooms farming, urban mushroom farming, and mushroom-livestock farming. In this study the condition of cultivation of several mushroom species and their edibility is included. Edible/medicinal mushrooms are well-known for their high content of many beneficial bioactive ingredients for human health such as antioxidants, ergosterols, lectins, phenolics/polyphenolics, polysaccharides, and terpenoids. These bioactives of mushrooms have the potential capability to treat and/or prevent several chronic diseases. The medicinal and prebiotic attributes of mushrooms still need more investigation, and this topic of “mushrooms in medicine” still has several open questions are needed to be answered in the future.

Keywords: Mushroom-worm farming, Mushroom-bee farming, Smart mushroom farming, Forest-mushrooms farming, Urban mushroom farming, Mushroom-livestock farming.

1. Introduction

Cultivation of mushrooms is thought to be back over four thousand years of many edible and medicinal mushroom species by the ancient Egyptians, Japan

and China (Cotter 2014). The cultivation of mushrooms needs an understanding of all development stages, which include the release of the spores (sporulation), their germination on a suitable

*Corresponding author e-mail: ramady2000@gmail.com

Received: 28/04/2023; Accepted: 08/05/2023

DOI: 10.21608/JENVBS.2023.207745.1216

©2023 National Information and Documentation Center (NIDOC)

growing media, colonization as much territory as possible to build up a competitive biomass, complete colonization and primordia formation, and mature mushrooms for producing spores, and the cycle starts again (Cotter 2014). Several reports have been published on mushroom cultivation from different points of view such as the cultivation of mushrooms as a sustainable integrated agriculture model (Ye et al. 2023), for mycoremediation (Sahithya et al. 2022), supporting selenium in removing mercury from soil (Pereira de Oliveira et al. 2023), for biofuel production (Leong et al. 2023), and biosynthesis of nanoparticles for removing organic pollutants (Chauhan et al. 2023).

Mushroom farming could be found in several types including mushrooms for healthy foods (Bell et al. 2022), mushroom-worm farming (Yang et al. 2023), mushroom-bee farming system, smart mushroom farming (Rahman et al. 2022), forest-mushrooms farming (Copena et al. 2022), urban mushroom farming (Dorr et al. 2021), and mushroom-livestock farming (Wang et al. 2022).

Edible mushrooms have unique nutritional attributes that allow use as a source of protein meat analog compared with animal meat and plant-based meat analogs (Wang and Zhao 2023). Many recent studies confirmed the bright side of mushrooms as a future generation healthy food (e.g., Okuda 2022; Bell et al. 2022). Many applications of biotechnology in mushroom farming, which focuses on traditional cultivation include mining biosynthetic gene clusters, precision breeding, developing mushroom chassis cells, and constructing cell factories for high value-added products (Zou et al. 2023). Several mushrooms have a great potential source of active metabolites and medicines (Bhambri et al. 2022).

Therefore, several therapeutic applications of mushroom including edible and medicinal ones were reported (Chugh et al. 2022; Ahmad et al. 2023), which may include more than 130 medicinal activities such as antioxidant, antitumor, cardioprotective and antiviral actions, immunomodulation, and radical scavenging (Chugh et al. 2022).

Therefore, this review presents an overview of the mushrooms and their cultivation and different antimicrobial and prebiotic properties, in addition, different mushroom farming types will be

highlighted.

2. Mushroom farming requirements

The cultivation of mushrooms is an important industry, which can gain a potential income. This industry can be carried out indoors or outdoors (in the open field or forests). Due to its importance, mushrooms have a promising strategy in the Nano-Food Lab (Debrecen university), which started several years ago (**Figure 1**). This plan was successfully translated into many publications and many patents in progress including many research areas such as cultivating edible mushroom in polluted soils (El-Ramady et al. 2021), the green biotechnology of mushrooms (El-Ramady et al. 2022a), the nutritional and medicinal attributes of edible mushrooms (El-Ramady et al. 2022b), the sustainable production of food and energy (El-Ramady et al. 2022c), the sustainable soil nano-management (Elsakhawy et al. 2022), sustainable applications in soil science (Fawzy et al. 2022), and edible mushrooms with focus on *Pleurotus* spp. (Törös et al. 2022), and *Lentinula* spp. (Hajdú et al. 2022).

Pleurotus mushrooms are globally well-cultivated on a large scale, accounting for 27% of their global production (Raman et al. 2021). Several mushrooms can adapt their growing to a wide range of temperatures, at relatively high humidity and high CO₂ levels without requiring specific controlled environmental conditions (Raman et al. 2021). In general, there are seven steps for mushroom cultivation are presented in **Figure 2**, whereas **Figure 3** includes some photos of mushroom farming (*Pleurotus ostreatus*). The main systems of mushroom cultivation could be the following:

I. Outdoor systems (e.g., logs, stumps, and wood chips), and

II. Indoor systems (e.g., bags under greenhouse conditions, bottles of king oyster, bags hung in a wall formation, horizontal shelf with bags, shelf cultivation of mushrooms, a-frame shelf with bags, tray cultivation of mushrooms, and sawdust blocks of mushrooms).

The main types of mushroom spawn include sawdust spawn, grain spawn, plug or dowel spawn, straw spawn, naturalized or wild spawn, and liquid spawn.

The substrate of cultivated mushrooms differs depending on the climate zone of the cultivation area and the system. For example, Cotter (2014) reported that the available substrate under temperate climate (16–29°C) may involve the following systems:

1- Logs and stumps (e.g., beefsteak, birch polypore, black poplar, cauliflower, and chicken of the woods),

2- Wood mulch or chips (e.g., brick top, king

Stropharia, and parasol),

3- Composts and livestock waste (e.g., almond portabella, and king Stropharia, parasol, shaggy mane, and composted livestock manure),

4- Agricultural waste, straw, plant debris (e.g., Elm oyster, king Stropharia, parasol, and shimeji),

5- Sawdust (e.g., black poplar, beefsteak, elm oyster, and hairy panus).

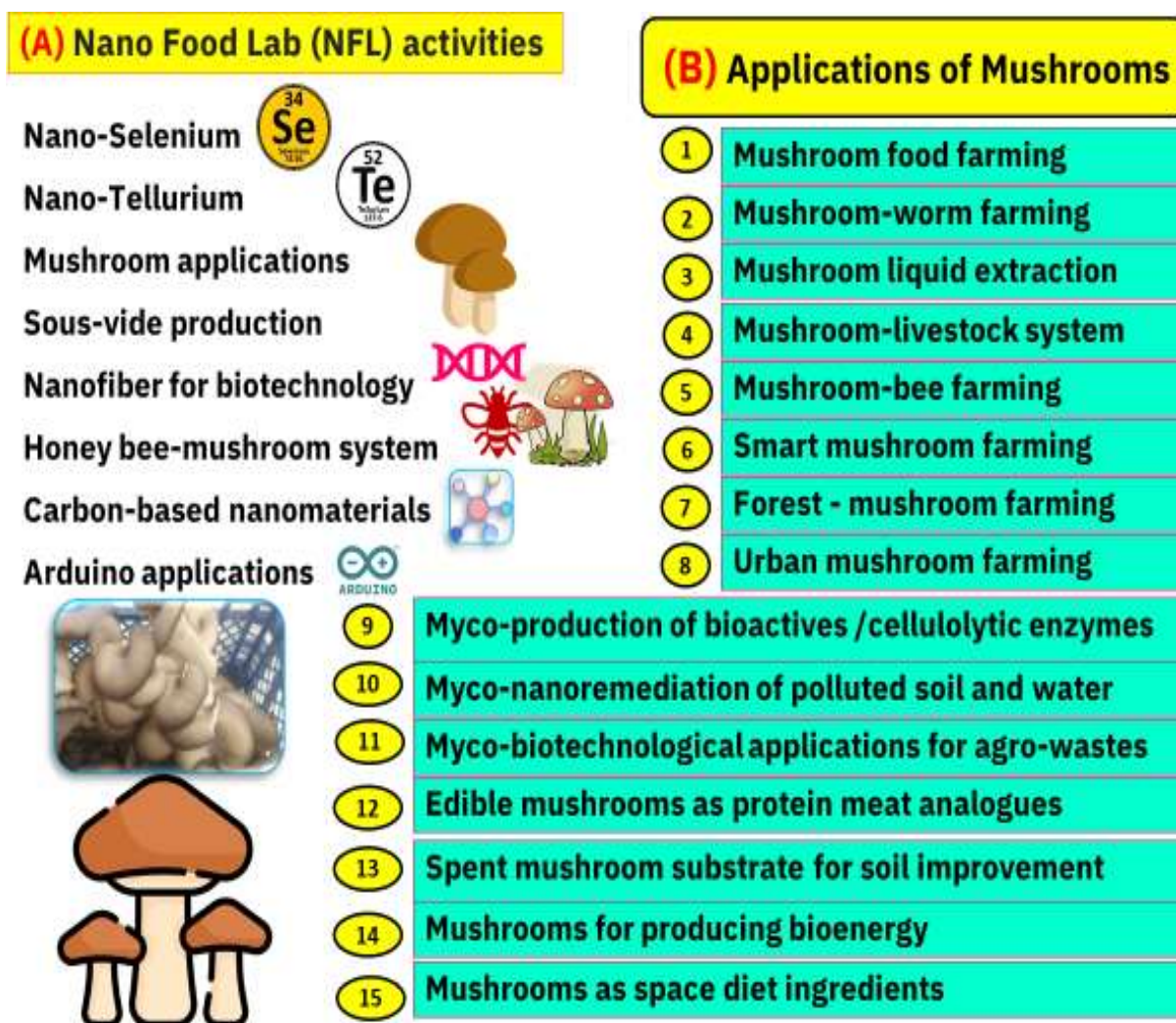


Fig. 1. List of activities of the Nano-Food Lab (Debrecen University, Hungary) in part (A), which includes many applications of mushrooms (part B).

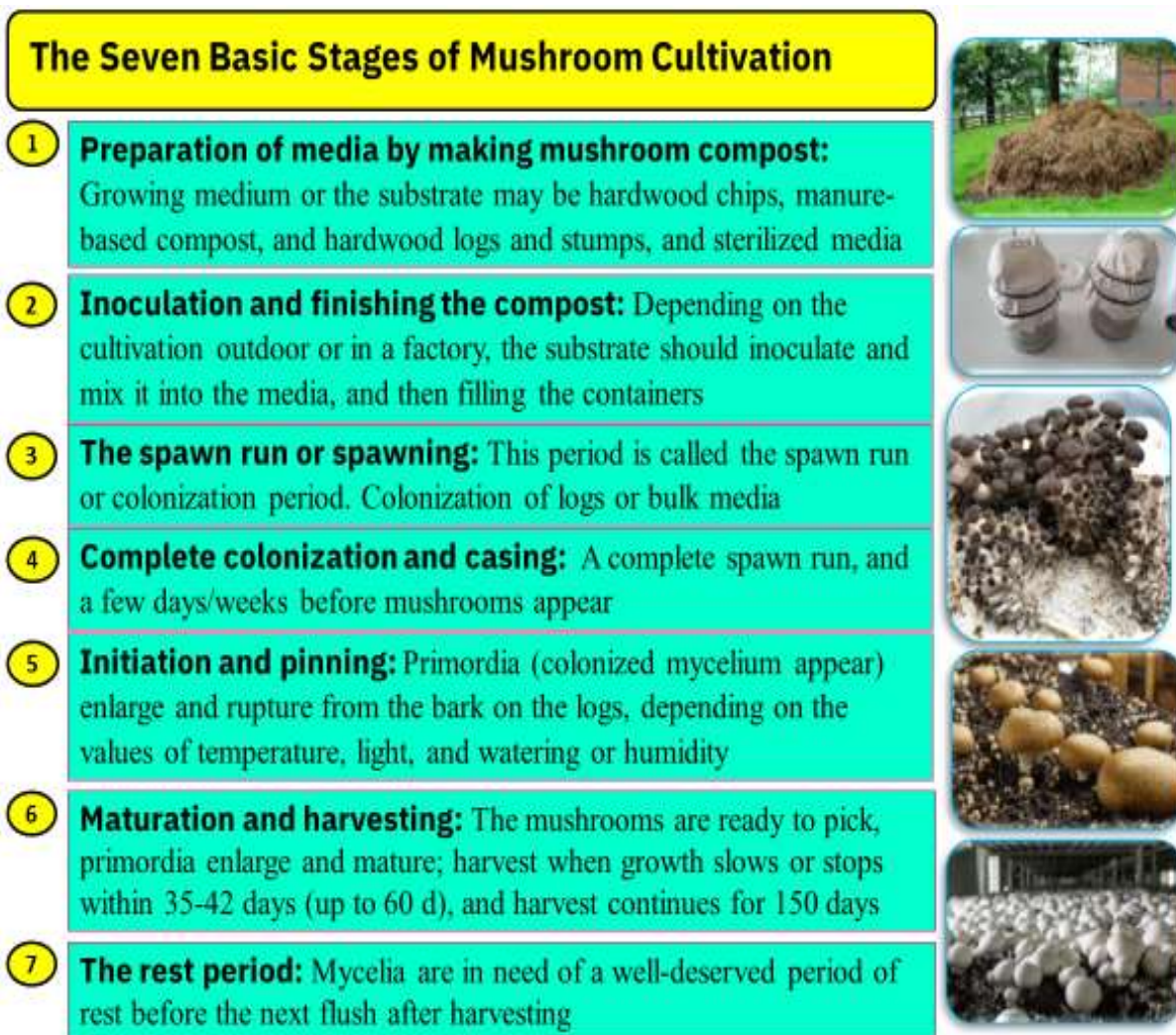
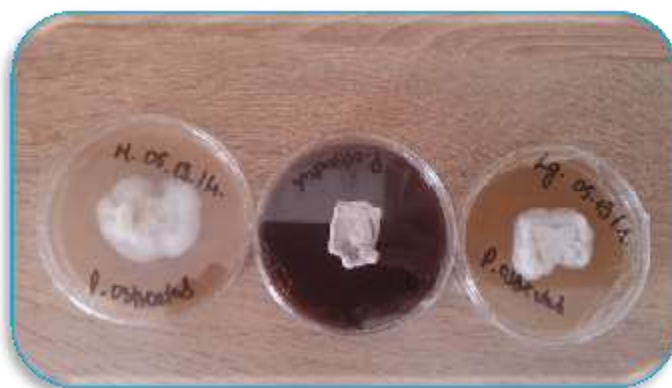


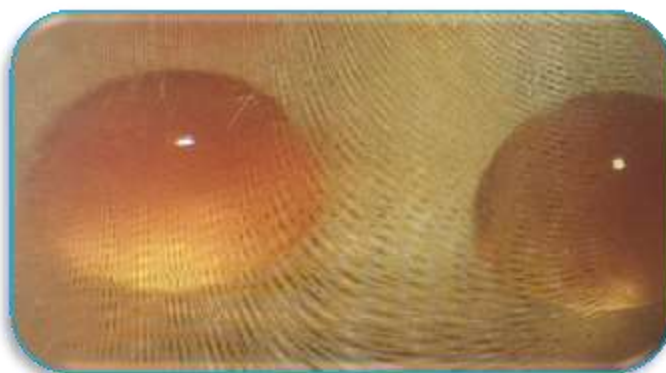
Fig. 2. The main steps in cultivation of mushrooms according to Cotter (2014).



Pleurotus ostreatus mycelium on different media (directly after vaccination; room temperature)



Pleurotus ostreatus mycelium on different media after 3 days from incubation stage at room temperature



The microscopic image of secondary metabolite products with orange ball appearance on oyster mycelium



The first harvest of *Pleurotus ostreatus* mushroom grown on corn husks based compost



The first harvest of fresh *Pleurotusostreatus* mushroom as a bouquet grown on corn husks-based compost



Freeze-dried *Pleurotusostreatus* mushrooms, which has been grown on corn husks-based compost



Fresh washed, sliced *Pleurotus ostreatus*

Fig. 3. Some main steps of oyster mushroom cultivation and harvesting from Nano Food Lab (Debrecen University, Hungary). All photos by GrétaTörös.

3. Mushroom farming types

The cultivation of mushrooms under certain conditions is called mushroom farming, and this activity could be performed with or without sharing other agricultural activities like forestry or livestock, the types of farming are presented in **Figure (4)**. Generally, wild mushrooms can grow (up to

10,000 different types of mushrooms), can be classified into 4 categories: parasitic, saprotrophic, mycorrhizal, and endophytic. The consumed mushroom substrate can be used in producing low-carbon biofuel (Leong et al. 2022). Although mushrooms have so many uses, the potential for producing a healthy food is quite important. It is very

important to highlight the edibility of many mushrooms in this study as presented in **Table 1**, and they are classified into edible, inedible, toxic, and unknown. The edibility and more growing details

besides English and scientific name of these groups of mushrooms are listed as well in Table 1.



Fig. 4. Mushroom farming systems including production of different farming types with focus on foods, livestock, vermicompost, etc.

Table 1. Some common mushrooms in the forestry system with its edibility (adapted from Yan 2023), and the scientific names according to <https://www.gbif.org/species>.

English name	Scientific name	Edibility and forest growing details
(1) Group of non-edible mushrooms cultivated in forests		
Coral Slime	<i>Ceratiomyxafruticulose</i> f. <i>aurantiaca</i> Jaap, (1922)	Rotten wood, sometimes on dead leaves. Not Edible
Scrambled Egg Slime	<i>Fuligoseptica</i> L. (F.H.Wigg., (1780)	Deciduous forests, on rotten wood, bark, or dead leaves. Not Edible
The Hidden Forest	<i>Stemonitisaxifera</i> (Bull.) T.Macbr., (1899)	Rotten Wood. Not Edible
Wolf's Milk Slime	<i>Lycogalaepidendrum</i> (L.) Fr., (1829)	Dead/Rotten wood. Not Edible.
Raspberry Slime Mold	<i>Tubiferaferruginosa</i> (Batsch) J.F.Gmel., (1792)	Dead wood, dead leaves, or humus. Not Edible
Lemon Drops or Lemon Disco	<i>Bisporellacitrina</i> (Batsch) Korf&S.E.Carp. (1974)	Dense colonies on dead branches, rotten wood. Prefers deciduous trees like beech. Not Edible
Canadian Round-headed Cordyceps	<i>Cordycepscanadensis</i> Ellis&Everh. (1898)	In humus under pines and deciduous trees. Latches on underground truffles, Not Edible
Adder's Tongue	<i>Cordycepsophioglossoides</i> (J.F.Gmel.) Link (1818)	Found in humus of deciduous and mixed forests (birch),latches on to underground truffles, Not Edible.
Black Knot of Cherry	<i>Dibotryonmorbosum</i> Schwein. Theiss.	Latches on to cherry trees such as

	& Syd. (1915)	<i>Prunusvirgiana</i> ; <i>Prunuspensylvanica</i> . Not Edible.
Agassiz's Lachnella	<i>Lachnellulaagassizii</i> (Berk. &M.A.Curtis) Dennis (1962)	In dense colonies on dead conifer bark, especially fir. Not edible.
Hairy Earth Tongue	<i>Trichoglossumhirsutum</i> (Pers.) Boud. (1907)	On humus or rotten wood. Not edible.
Dead Man's Fingers	<i>Xylariapolymorpha</i> (Pers.) Grev. (1824)	On dead branches, trunks, or logs. Even on live trees, especially on deciduous trees such as beech. Not edible.
Black Witche's Butter	<i>Exidiaglandulosa</i> (Bull.) Fr. (1822)	On dead branches from deciduous trees. Not edible.
White Coral Jelly Fungus	<i>Tremellareticulata</i> (Berk.) Farl. (1908)	On ground or on rotten wood or leaves. Especially in maple/oak forests. Not edible.
Perennial Polypore	<i>Coltriciaperennis</i> (L.) Murrill (1903)	On the ground in disturbed forests such as clear cuts or wildfire mainly in coniferous forests with birch and aspen. Not Edible.
Woolyvelvet polypore or Velvet Rosette	<i>Onniatomentosa</i> (Fr.) P.Karst. (1889)	On buried roots in coniferous forests. Not Edible.
Winter Polypore	<i>Polyporusbrumalis</i> (Pers.) Fr. (1818)	On dead wood, especially birch wood. Not edible.
Variegated Polypore	<i>Polyporusvarius</i> (Pers.) Fr. (1821)	On dead wood in deciduous forests. Not Edible.
Reddish Brown Crust	<i>Hymenochaetetabacina</i> (Sowerby) Lév. (1846)	On dead branches and trunks, in deciduous forests. Not edible.
Red Tree Brain	<i>Peniophorarufa</i> (Fr.) Boidin (1958)	In dense colonies, on dead branches, trunks of quaking aspen, or willow. Not Edible.
Reddish Brown Crust	<i>Hymenochaetetabacina</i> (Sowerby) Lév. (1846)	On dead branches and trunks, in deciduous forests. Not edible.
Hairy Parchement	<i>Stereumhirsutum</i> (Willd.) Pers. (1800)	On dead wood of deciduous trees, such as oak, maple, and beech. Not edible.
Milk-white Toothed Polypore	<i>Irpexlacteus</i> (Fr.), Fr. (1828)	On dead branches and trunk of deciduous trees. Not Edible.
Tin Maze Flat Polypore	<i>Daedaleopsisconfragosa</i> (Bolton) J.Schröt. (1888)	On dead trunks of deciduous trees, mainly on beech, birch, willows,alders. Not Edible.
Mossy Maze Polypore	<i>Cerrenaunicolor</i> (Bull.) Murrill (1903)	In large colonies on dead wood, or even living deciduous trees. Not edible.
Split-gilled Bracket	<i>Schizophyllum commune</i> Fr. (1815)	Dead branches of deciduous trees. Not Edible.
Rusty-gilled Polypore	<i>Gloeophyllumsepiarium</i> (Wulfen) P.Karst. (1882)	On dead trunks, or logs of coniferous trees. Not edible.
Timber Polypore	<i>Fomesfomentarius</i> (L.), Fr. (1849)	On trunks of living or dead deciduous trees, such as birch. Not edible.
Red-belted Polypore/ Red-Banded Polypore	<i>Fomitopsispinicola</i> (Sw.) P.Karst. (1881)	On dead trunks, mostly on conifers. Not Edible.
Artist's Conk	<i>Ganodermaapplanatum</i> (Pers.) Pat. (1887)	On trunks of dead deciduous trees. Not edible.
Resinous Polypore	<i>Ischnodermaresinosum</i> (Schrad.) P.Karst. (1879)	On logs, and trunk of dead, deciduous trees. Not edible.
Scaly Polypore	<i>Polyporusquamosus</i> (Huds.) Quélet (1886)	On trunks and logs of wounded deciduous trees. Not Edible.
False Timber Polypore	<i>Phellinus cinereus</i> Niemelä	On stumps, mostly of birch trees. Not edible.
Birch Polypore	<i>Piptoporusbetulinus</i> (Bull.) P.Karst. (1881)	On dead birch trees. Not edible.
Turkey Tail Polypore	<i>Trametesversicolor</i> (L.) Lloyd (1921)	On stumps, trunks, logs, or branches of deciduous trees. Not edible, but Medicinal.
Cinnabar-red Plypore (Cinnabar Bracket)	<i>Pycnoporuscinnabarinus</i> (Jacq.) P.Karst. (1881)	On stumps, logs, or trunks of deciduous trees, commonly on cherry. Not edible.
Parchment Bracket or Violet-toothed polypore	<i>Trichaptumbiforme</i> (Fr.) Ryvarden (1972)	On stumps, trunks, and logs of deciduous trees, sometimes covering the whole surface.

Northern Tooth	<i>Climacodonseptentrionalis</i> (Fr.) P.Karst. (1881)	Not edible. On trunks of dead or living deciduous trees such as elm and maple. Not edible.
Fragrant Hydnum (SweetgrassHydnellum)	<i>Hydnellumsuaveolens</i> (Scop.) P.Karst. (1879)	On mossy forest floor, lichen or needle covered soil. Not edible.
Flat Crep	<i>Crepidotusapplanatus</i> (Pers.) P.Kumm. (1871)	Dispersed on dead wood in deciduous forests. Not edible.
Bear Lentinus	<i>Lentinellusursinus</i> (Fr.) Kühner, (1926)	On dead wood in deciduous forests. Not edible.
Luminescent Panellus	<i>Panellusstypticus</i> (Bull.) P.Karst. (1879)	In bunches, on rotten wood of deciduous forests. Not edible.
Orange Mock Oyster	<i>Phyllotopsisnidulans</i> (Pers.) Singer (1936)	In bunches, on dead wood of coniferous and deciduous forests. Not edible.
Pinwheel Marasmius	<i>Marasmiusrotula</i> (Scop.) Fr. (1838)	In bunches on rotten wood, or on dead leaf litter, or within stacks of twigs in deciduous forests. Not edible.
Bleeding Mycena or Bleeding Fairy Helmet	<i>Mycenahaematopus</i> (Pers.) P.Kumm. (1871)	In bunches on rotten wood of deciduous trees. Stem bleeds a blood-colored latex. Not Edible.
Wrinkled Mycena	<i>Mycenagalericulata</i> (Scop.) Gray (1821)	In bunches on rotten wood of deciduous trees. Not edible.
Pink Mycena	<i>Mycenapura</i> -(Pers.) P. Kumm. (1871)	On humus in coniferous, mixed, or deciduous forests. Not Edible.
Orange Mycena	<i>Mycenaleaiana</i> (Berk.) Sacc. (1891)	In bunches on dead logs of deciduous trees. Not edible.
Fuzzy Foot	<i>Xeromphalinacampanella</i> (Batsch) Kühner&Maire (1953)	In dense clusters on stumps, and trunks of rotten coniferous trees. Not edible.
Scaly Stropharia	<i>Strophariasquamosa</i> var. <i>thrausta</i> (Kalchbr.) Masee	On buried wood, in forests or fields. Not Edible.
(2) Group of edible mushrooms cultivated in forests		
Orange PeelFungus	<i>Aleuriaaurantia</i> (Pers.) Fuckel	Bare, usually sandy soil. Edible
False Morel	<i>Giromitraesculenta</i> (Pers. ex Pers.) Fr. (1849)	Open, sandy soils, in coniferous forests, particularly white pine, and mixed forests. Edible for some, but not recommended.
Lobster Mushroom	<i>Hypomyceslactiflorum</i> Schwein.,Tul. &C.Tul. (1860)	Latches on to russula, and lactarius mushrooms. In coniferous or mixed forests. Edible, with great texture.
Common Jelly Baby	<i>Leotialubrica</i>	In dense colonies on rotten wood/ directly on the ground. Edible, but glutinous.
Black Morel	<i>Morchellaelata</i> Pers. Fr. (1822)	Deciduous forests, especially under poplar, but also in mixed forests. Edible, tasty, but better blanch them first.
Yellow Morel	<i>Morchellaesculenta</i> (L.) Pers. (1801)	Deciduous forests, especially under elms, and poplars. Edible, one of the tastiest, but better blanch them first.
Irregular Mitrula or Irregular Earth Tongue Bay Peziza or bay cup	<i>Neolectairregularis</i> (Peck) Korf&J.K.Rogers (1971) <i>Pezizabadia</i> Pers. (1800)	On moss, or forest litter such as needles, in coniferous or mixed forests. Edible. In deciduous or mixed forests, often found on sandy soil. Edible.
Scarlet Cup	<i>Sarcoscyphaaustriaca</i> (Beck ex Sacc.) Boud. (1907)	On the soil, in burrowed wood, in deciduous forests such as maple forests. Edible.
Fairy Butter	<i>Dacrymyceschrysospermus</i> Berk. &M.A.Curtis (1873)	On dead wood of conifers. Edible.
Tree Ear	<i>Auriculariaauricula-judae</i> (Bull.) Qué. (1886)	On dead wood, especially on conifers like fir and spruce. Edible.
Toothed Jelly Fungus	<i>Pseudohydnumgelatinosum</i> (Scop.) P.Karst. (1868)	On rotten wood in coniferous forests. Edible, but not particularly tasty.
Fake Coral Fungus	<i>Tremellodendron pallidum</i> Burt. (1915)	On ground in deciduous or mixed forests. Edible.

Apricot Jelly Fungus	<i>Tremiscushelvelloides</i> (DC.) Donk (1958)	On ground, or rotten wood in coniferous or mixed forests. Edible, but more decorative than tasty.
Sheep Polypore	<i>Albatrellusovinus</i> (Schaeff.) Kotl. &Pouzar (1957)	Under fir or spruce forests. Edible, quite tasty.
Chaga	<i>Inonotus obliquus</i> (Fr.) Pilát (1942)	Latches on exclusively on birch trees. Highly medicinal, can be used as coffee alternative. Edible
Chicken of the Woods	<i>Laetiporussulphureus</i> (Bull.) Murrill (1920)	On trunks and logs of deciduous trees. Edible, but with variety of results.
Comb Tooth	<i>Hericiumcoralloides</i> (Scop.) Pers. (1794)	On trunk of dead or living deciduous trees. Edible.
Sweet Tooth (Wood Hedgehog)	<i>Hydnumrepandum</i> L. (1753)	In mixed, deciduous or coniferous forests. Edible, very tasty.
Fairy Fingers	<i>Clavariafragilis</i> Holmsk. (1790)	In dense groups on the ground within herbs, and humus, in deciduous forests. Edible.
Purple coral, or the purple fairy club	<i>Clavariapurpurea</i> (Fr.) Dentinger&D.J.McLaughlin (2007)	In colonies on the ground, within herbs, or humus, in coniferous forests. Edible.
Whit coral fungus or the crested coral fungus	<i>Clavulinacristata</i> (Holmsk.) J. Schröt. (1888)	On the ground within moss, and humus, in coniferous or mixed forests. Edible.
Crown-tipped Coral or Candelabra Coral	<i>Clavicornapyxidata</i> (Pers.) Doty (1947)	On rotten wood of deciduous trees, mainly willow, or aspen. Edible, but mediocre.
Spindle-shaped Clavaria	<i>Clavulinopsis fusiformis</i> (Sowerby) Corner (1950)	In dense clusters within herbs, or on bare soil, within coniferous or mixed forests. Edible.
Chanterelle	<i>Cantharelluscibarius</i> Fr. (1821)	In coniferous, and mixed forests, and more rarely in deciduous forests. Edible, tasty.
Appalachian Chanterelle	<i>Cantharellusappalachiensis</i> R.H. Petersen (1971)	In deciduous forests, often under oaks, and beech. Edible, tasty.
Cinnabar Chanterelle	<i>Cantharelluscinnabarinus</i> (Schwein.) Schwein. (1832)	On the forest floor, within moss, or along paths, in deciduous forests, often under oaks. Edible, tasty.
Black Trumpet	<i>Craterellusfallax</i> A.H.Sm. (1968)	In deciduous, and mixed forests. Edible, tasty.
Trumpet Chanterelle or Yellowfoot	<i>Craterellustubaeformis</i> (Fr.) Quél. (1888)	In coniferous, or mixed forests. In moist areas such as peat bogs, in sphagnum moss. Edible, tasty.
Oyster Mushroom	<i>Pleurotostreatatus</i> (Jacq. ex Fr.) P.Kumm. (1871)	In bunches, on living or dead deciduous trees, especially on maple, oak, beech, and birch. Edible, tasty.
Angel's Wings	<i>Pleurocybellaporrigens</i> (Pers.) Singer (1947)	In bunches, on rotten wood of conifers. Edible, tasty.
Summer or pale Oyster	<i>Pleurotuspulmonarius</i> (Fr.) Quél. (1872)	In bunches, on living or dead deciduous trees, such as maple, beech, oaks, and birch. Edible, tasty.
Late Fall Oyster	<i>Sarcomyxaserotina</i> (Pers.) P. Karst. (1891)	In bunches, on dead deciduous trees. Edible, tasty.
Golden Waxy Cap	<i>Hygrocybechlorophana</i> (Fr.) Wünsche (1877)	In coniferous, mixed, or deciduous forests. Edible, but not recommended.
Chanterelle or Goblet Waxy Cap	<i>Hygrocybecantharellus</i> (Schwein.) Murrill (1911)	On extremely rotten wood covered in moss, and on sphagnum moss within humid peat bogs. Edible
Scarlet Waxy Cap	<i>Hygrocybecoccinea</i> (Schaeff.) P. Kumm. (1871)	On the forest floor in deciduous, and mixed forests. Edible.
Witch's Hat or Blackening Waxcap	<i>Hygrocybeconica</i> (Schaeff.) P.Kumm. (1871)	On the forest floor in deciduous, mixed, or coniferous forests; or in open meadows, and groves. Edible, but not recommended.
Virginal Hygrophorus	<i>Hygrocybevirginea</i> (Wulfen) P.D.Orton& Watling (1969)	On poorly drained soil, sometimes withing sphagnum, in coniferous, and mixed forests,

Larch Waxy Cap	<i>Hygrophorus speciosus</i> Peck, (1878)	in pastures, and meadows. Edible. Associates with larch trees in humid, coniferous forests. Found in peat bogs, and in sphagnum. Edible.
Dark Honey Fungus	<i>Armillaria ostoyae</i> (Romagnesi) Herink (1973)	In dense bunches on tree stumps, roots, or dead trunks in deciduous or coniferous forests. Edible.
Grayling or the Humpback	<i>Cantharellula umbonata</i> (J.F.Gmel.) Singer (1936)	On mossy forest floors, in meadows, or groves of coniferous or mixed forests. Edible, tasty.
Club Foot	<i>Clitocybe clavipes</i> (Pers.) P.Kumm. (1871)	Mostly in coniferous, but sometimes in deciduous forests. Edible, but mediocre. Not to mix with alcohol.
Funnel Clitocybe	<i>Clitocybe gibba</i> (Pers.) P. Kumm. (1871)	On dead leaves, in deciduous or mixed forests. Edible, tasty.
Anis-scented Clitocybe	<i>Clitocybe odora</i> (Bull.) P.Kumm. (1871)	On leaf litter in deciduous or mixed forests. Edible, and aromatic.
False Chanterelle	<i>Hygrophoropsis aurantiaca</i> (Wulfen) Maire ex Martin-Sans (1921)	In little groups on the ground or on rotten wood in mixed or coniferous forests. Edible, tasty.
Pungent Cystoderma	<i>Cystoderma amianthinum</i> var. <i>rugosoreticulatum</i> (F.Lorinser) Bon (1999)	On forest floor covered with moss, and conifer needles. Edible, but not recommended.
Bicolored Laccaria	<i>Laccaria bicolor</i> (Maire) P.D.Orton (1960)	On the floor, or within moss under conifers, especially pine. Edible.
Iris-scented Lepista	<i>Lepista irina</i> (Fr.) H.E.Bigelow (1959)	In deciduous, mixed, or coniferous forests, within herbs on open ground. Edible, may cause gastrointestinal problems.
Fetid Tricholoma or booted knight	<i>Tricholoma focale</i> (Fr.) Ricken (1914)	In coniferous forests, as well as deciduous, and mixed. Edible, but not great.
Canary Trich or Yellow Knight	<i>Tricholoma equestre</i> (L.) P.Kumm. (1871)	In coniferous or mixed forests, often under pine trees. Edible, but eat in small quantity
Shingled Trich	<i>Tricholoma imbricatum</i> (Fr.) P.Kumm. (1871)	In bunches, within coniferous, or mixed forests, especially near jackpine. Edible, but eat in small quantities.
Sticky Gray Trich or Coalman	<i>Tricholoma portentosum</i> (Fr.) Quéf. (1873)	In coniferous forests, but sometimes under beech trees. Edible, tasty.
Soapy Trich	<i>Tricholoma saponaceum</i> (Fr.) P.Kumm. (1871)	In coniferous forests, often under spruce, but also in deciduous or mixed forests. Edible, but not recommended.
Brownish-yellow Tricholoma	<i>Tricholoma tramsmutans</i> (Peck) Sacc. (1887)	In coniferous, or mixed forests, often under birch trees, or growing alongside moss. Edible, but mediocre.
Fibril Trich	<i>Tricholoma virgatum</i> (Fr.) P.Kumm. (1871)	In coniferous, or mixed forests, often growing alongside moss. Edible (mediocre)
Russet-scaly Trich	<i>Tricholoma vaccinum</i> (Schaeff.) P.Kumm. (1871)	In coniferous or mixed forests. Edible, but mediocre.
Oak-loving Collybia	<i>Gymnopus dryophilus</i> (Bull.) Murrill (1916)	On humus in deciduous, mixed, coniferous forests, especially near oak trees. Edible.
Lawn Mower's Mushrooms	<i>Panaeolus foenisecii</i> (Pers.) J.Schröt. (1926)	On lawns, and in prairies. Edible, but contains low amounts of psilocybin.
(3) Group of toxic mushrooms		
Eastern American Jack O'Lantern	<i>Omphalotus illudens</i> (Schwein.) Bresinsky & Besl	In bunches on trunks, or roots of oak trees. Extremely toxic.
Red-brown TrichorTacked Knight	<i>Tricholoma pessundatum</i> (Fr.) Quéf. (1872)	In coniferous forests. Not edible, toxic.
Red-gilled Cort or Surprise Webcap	<i>Cortinarius semisanguineus</i> (Fr.) Gillet (1876)	In coniferous, mixed, or oak forests. Not edible, dangerous.
Poison Pie	<i>Hebeloma crustuliniforme</i> (Bull. ex	In deciduous, or coniferous forests. Also on

Straw-colored Fiber Head or Split Fibrecap	St. Amans.) Quél. <i>Inocyberimos</i> (Bull.) P. Kumm. (1871)	lawns, and other open areas. Toxic. In deciduous, mixed, or coniferous forests. Toxic.
White Fiber Head	<i>Inocybegeophylla</i> (Bull.) P. Kumm. (1871)	On the ground in deciduous, mixed, or coniferous forests. Toxic.
Marginate Galerina	<i>Galerinamarginata</i> (Batsch) Kühner (1935)	In bunches on trunk of rotten deciduous, and coniferous trees. Toxic, DEADLY.
Laughing Mushroom or Yellow Gymnopilus	<i>Gymnopilusluteus</i> (Peck) Hesler (1969)	In dense bunches, on trunk and logs of dead deciduous, and coniferous trees. Not edible, contains hallucinogenic toxins.
Shaggy-stalked Lepiota	<i>Lepiotaclypeolaria</i> (Bull.) P. Kumm. (1871)	In coniferous or mixed forests. Toxic.
Rosy Entoloma or Wood Pinkgill	<i>Entolomarhodopolium</i> (Fr.) P. Kumm. (1871)	On forest floors. Toxic.
Lead Poisoner or Livid Pinkgill	<i>Entolomasinuatum</i> (Bull.) P. Kumm. (1871)	On forest floors. Highly Toxic.
Destroying Angel	<i>Amanita virosa</i> Bertill. (1866)	In mixed forests, especially under birch. Toxic, Deadly.
Poison Paxillus	<i>Paxillus involutus</i> (Batsch) Fr. (1838)	On forest floors of coniferous, mixed or deciduous forests. Toxic.
Common Scleroderma or common earthball	<i>Scleroderma citrinum</i> Pers. (1801)	Near stumps, rotten wood, in forests or open grounds. Not edible, toxic.
(4) Group of unknown edibility mushrooms		
Ocellate Collybia	<i>Clitocybula oculus</i> (Peck) Singer (1962)	On trunks, and logs of dead deciduous trees, often covering large surfaces. Unknown.
Smear Cort	<i>Cortinariustrivialis</i> J.E. Lange (1940)	In deciduous, coniferous, or mixed forests. Unknown.
Variable Cort	<i>Cortinarius multififormis</i> Fr. (1838)	In mixed, or coniferous forests. Unknown.
Pungeant Cort or Gassy Webcap	<i>Cortinariustraganus</i> (Fr.) Fr. (1838)	In mixed, or coniferous forests. Unknown.
Violet Cort or Violet Webcap	<i>Cortinarius violaceus</i> (L.) Gray (1821)	In mixed, or coniferous forests, often near birch trees. Unknown.
Maple Agrocybe	<i>Agrocybe acericola</i> (Peck) Singer (1950)	On wood chips, and rotten wood of deciduous trees. Unknown.
Lemon-yellow Pholiota	<i>Pholiotalimonella</i> (Peck) Sacc. (1887)	In bunches on rotten wood and wounds of deciduous trees. Unknown.
Dark-centered Lactarius or Eye Spot Milky	<i>Lactarius oculatus</i> (Peck) Burl. (1907)	In small bunches on humid soils, along moss in coniferous or mixed forests. Unknown.
Common Lactarius or Tacked Milkcap	<i>Lactariustrivialis</i> (Fr.), Fr. (1838)	In coniferous forests. Unknown.

4. Mushroom antimicrobial and prebiotic attributes

No doubt that edible mushrooms can be considered an important source for healthy food for the next-generation. These edible mushrooms have a great nutritional value due to their rich in ash (7-17%), dietary fiber (16-20%), proteins (30-48%), fat (1-4%), carbohydrate (12.5-40%), minerals, and vitamins like B₁, B₂, B₁₂, C, D, and E (Raman et al. 2021; Bhambri et al. 2022), as well as other bioactive components including alkaloids, lactones, polyphenolic compounds, polysaccharides, sesquiterpenes, sterols, and terpenoids (El-Ramady et al. 2022b). These bioactive ingredients are considered health-promoting supplements when extracted from edible mushrooms and applied to human foods. Several human diseases have been treated using edible mushroom extracts due to their biological impacts particularly antidiabetic,

anticancer, hepatoprotective, antiviral, antioxidant, immune-potentiating, and hypo-cholesterol impacts (Chugh et al. 2022). The health benefits of some common mushrooms were listed in **Figure (5)**. Mushroom protein can be considered a novel protein alternative (Ayimbila and Keawsompong 2023). Concerning the prebiotic properties of mushrooms, they exhibited distinguished influence as prebiotic properties due to their high content of many prebiotic components (e.g., chitin, hemicellulose, β and α -glucans, mannans, xylans, galactans, and inulin) as reported by many studies (e.g., Jayachandran et al. 2017; Moumita and Das 2022; Zhang et al. 2022). The prebiotic action of mushrooms may express as a stimulator of the growth of gut microbiota, conferring health benefits to the host.



Horse Mushroom

Agaricusarvensis L. (Schaeff.) 1774

Uses: cancer, cardiovascular disease, immune diseases, lower back pain



Champignon, Button Mushroom, Portobello

Agaricusbisporus L. (J.E.Lange) Imbach (1946)

Cancer, antioxidant, antimicrobial, cognitive function, cardiovascular disease, age, gut health



Field Mushroom

Agaricuscampestris L.

Diabetes, cancer, antimicrobial, antioxidant, lung cancer, fatigue



Almond Mushroom

*Agaricussubrufescens*L. Peck (1893)

Cancer, allergies, diabetes, dermatitis, hepatitis, infections, tumors, inflammation, high cholesterol



Jelly Ear Fungus

Auriculariaauricula-judae(Bull.) Quél. (1886)

Inflammation, sore throat, fever, healthy blood, antioxidant,tumor, anticoagulant



Turkey Tail

Coriolus versicolor L. (Quél.) (1886)

Immune System, cancer, diabetes



Enokitake, Enoki

Flammulina velutipes (Curtis) Singer (1951)

Cancer, immune system, antimicrobial, antioxidant, neurodegenerative diseases, high cholesterol, inflammation, aging



Birch Polypore

Fomitopsis betulina (Bull.) B.K. Cui et al. (2016)

Antibiotic, purgative, inflammation, viruses, styptic, antiseptic, cancer, HIV



Agarikon

Fomitopsis officinalis (Vill.) (1941)

Pulmonary diseases, rheumatism, asthma, viruses, antibacterial, tuberculosis



Red-Banded Polypore

Fomitopsis spicicola (Sw.) P. Karst. (1881)

Headaches, laxative, inflammation, antimicrobial, styptic



Reishi

Ganoderma lucidum (Curtis) P. Karst. (1881)

Inflammation, cancer, respiratory issues, asthma, insomnia, arthritis, allergies



Maitake, Hen of the woods

Grifola frondosa (Dicks.) Gray (1821)

Cancer, diabetes, tumors



Lion's Mane

Hericium erinaceus (Bull.) Persoon (1797)

Memory, cognitive health, strength, vigor digestion, inflammation, Alzheimer's, anxiety



Chicken of the Woods

Laetiporus sulphureus

Cancer, hypoglycemia, inflammation, antioxidant, antimicrobial, high cholesterol, anticoagulant



Devil's Tooth

Hydnellum peckii Banker (1912)

Anticoagulant, Antibacterial



Chaga

Inonotus obliquus (Ach. ex Pers.) Pilát (1942)

Cancer, tumors, wounds, swelling, diabetes, antioxidant, antimicrobial, HIV, Hepatitis C



Shiitake

Lentinula edodes (Berk.) Pegler (1976)

High cholesterol, antimicrobial, tumor, immune system, cancer



Tiger's Milk

Lignosus rhinoceros (Cooke) Ryvarden (1972)

Neurodegenerative diseases, fever, Itching, asthma, Cancer



Chinese Caterpillar Fungus

Ophiocordyceps sinensis (Berk.) (2007)

General health, endurance, stamina, immune issues, fatigue, Progesterone, kidney function



Bitter Oyster Mushroom

Panellus stipiticus (Bull.) P.Karst. (1879)

Hemorrhaging



Black Hoof Mushroom

Phellinus linteus (Berk. MACurtis) Teng (1963)

Cancer, menstruation & gastrointestinal issues, antioxidant, diabetes, antimicrobial, viruses, Inflammation, Myocardial ischemia reperfusion



Oyster Mushroom

Pleurotus ostreatus (Jacq.) P. Kumm. 1871

Diabetes, hyperlipidemia, cancer, infections, high cholesterol, fungal diseases, tumors, antioxidant, anti-aging



Horn of Plenty

Pleurotus cornucopiae (Paulet) Rolland (1910)

High Blood Pressure



Kingtuber mushroom

Pleurotus tuber-regium (Rumph. ex Fr.) Singer 1951

Cold, fever

**Abalone***Pleurotus cystidiosus* O.K. Mill. (1969)

Diabetes

**Matsutake***Tricholoma matsutake*

Antimicrobial, Anti-inflammatory

**Stout Camphor Fungus***Taiwanofungus camphoratus* Wu et al. (2004)Cancer, Allergies, Fatigue, Liver Issues,
Antioxidant, Diabetes, Hepatitis B**NOT available English name***Tolyptocladium inflatum* W. Gams (1971)Immunosuppressant, inflammation, fungal diseases,
Psoriasis, Eczema, Crohn's Disease, Diabetes

Fig. 5. Some common mushrooms and their medical use <https://www.gbif.org/species> and photos from <https://www.pexels.com/> and Wikipedia accessed on 18.04.2023.

According to Bhambri et al. (2022), the suggested mechanism of such medicinal mushrooms presented as follows:

1- Anti-cancer of *Agaricus* spp. by inhibiting cell proliferation of some cancer cell lines, antioxidant activities, and anti-inflammatory due to many metabolite components such as phenolic, sterols, indole compounds and nutraceuticals (Usman et al. 2021),

2- Antitumor, antioxidative, hypolipidemic, and antibacterial effects of *Coprinus* spp. by regulating the blood glucose level, hypoglycemic and antioxidative homeostasis due to forming carbohydrates, dietary fibers, and phenolic compounds (Stilinic et al. 2020),

3- Anti-inflammatory, antitumor activity against both ascites as well as solid tumors of ethanolic extracts, and high antioxidant activity of *Morchella* spp. by increasing the cytotoxic effect and as immunomodulator because of organic acids, free fatty acids,

flavonoids, triglycerides, and sterols (Dissanayake et al. 2021),

4- Antioxidant, antihyperglycemic, antimicrobial, iron-chelation, wound healing, cytotoxicity, anti-hypoxic, anti-acid inflammatory of *Cortinarius* spp. by inhibiting protein synthesis due to presence of amino acids, and orellanine (Meena et al. 2020), and

5- Antitumor, anti-inflammation, antiviral, antidiabetic, antioxidation, anti-hypertensive, immune-enhancing, immunomodulation, hyperlipidemia and hyperglycemia by *Grifola* spp. as an immunomodulator by the action of glucans, sesquiterpenes, and glycoproteins (Su et al. 2020).

Is the world of mushroom a real treasure for the scientific research? The answer simply is yes. This can explain in only one sentence that several mushrooms have a great potential as nutritional, pharmaceutical, and medicinal impacts. Regarding the nutritional value of mushrooms, they have enough and high values of many essential nutritional

compounds like fiber, protein, carbohydrates and vitamins. Concerning the pharmaceutical attributes of mushrooms, they contain several metabolite compounds that can exhibit the pharmaceutical behavior in human like antifungal, antibacterial, antiviral, and tumor attenuating. Mushrooms are promising prebiotic agents due to their stimulation of the growth of gut microbiota, which confer health benefits to the host.

5. Conclusions

It could conclude that mushrooms have a distinguished nutritional value, due to their enough and high values of many essential nutritious compounds like protein, fiber, carbohydrates and vitamins. Mushrooms also have pharmaceutical and medicinal attributes, because of them contain many metabolite compounds, which can exhibit the pharmaceutical behavior in human like antifungal, antibacterial, antiviral, and tumor attenuating. Mushrooms are promising prebiotic agents due to their stimulation of the growth of gut microbiota, which confer health benefits to the host. Therefore, it is expected that enormous of research are needed to be carried out on different edible and medical mushrooms to discover more and more benefits of this treasure.

Ethics approval and consent to participate: This article does not contain any studies with human participants or animals performed by any of the authors.

Consent for publication: All authors declare their consent for publication.

Conflicts of Interest: The author declares no conflict of interest.

Contribution of Authors: All authors shared in writing, editing and revising the MS and agree to its publication.

Acknowledgments: G. Törös and the authors thanks the 2020-1.1.2-PIACI-KFI-2020-00100 Project “Development of innovative food raw materials based on Maillard reaction by functional transformation of traditional and exotic mushrooms for food and medicinal purposes” for financializing and supporting this work.

6. References

Ahmad I, Arif M, Xu M, Zhang J, Ding Y, Lyu F (2023). Therapeutic values and nutraceutical properties of shiitake mushroom (*Lentinula edodes*): A review. Trends in Food Science & Technology, 134, 123-135. <https://doi.org/10.1016/j.tifs.2023.03.007>.

Ayimbila F, Keawsompong S (2023). Nutritional Quality and Biological Application of Mushroom Protein as a Novel Protein Alternative. Current nutrition reports, 1–

18. Advance online publication. <https://doi.org/10.1007/s13668-023-00468-x>

Bell V, Silva CRPG, Guina J and Fernandes TH (2022). Mushrooms as future generation healthy foods. Front. Nutr. 9:1050099. Doi: 10.3389/fnut.2022.1050099

Bhambri A, Srivastava M, Mahale VG, Mahale S and Karn SK (2022). Mushrooms as Potential Sources of Active Metabolites and Medicines. Front. Microbiol. 13:837266. Doi: 10.3389/fmicb.2022.837266

Bray R (2020). Medicinal Mushrooms: A Practical Guide to Healing Mushrooms (Urban Homesteading), Independently published

Chauhan N, Thakur N, Kumari A, Khatana C, Sharma R (2023). Mushroom and silk sericin extract mediated ZnO nanoparticles for removal of organic pollutants and microorganisms. South African Journal of Botany, 153, 370-381. <https://doi.org/10.1016/j.sajb.2023.01.001>.

Chugh RM, Mittal P, MP N, Arora T, Bhattacharya T, Chopra H, Cavalu S and Gautam RK (2022). Fungal Mushrooms: A Natural Compound With Therapeutic Applications. Front. Pharmacol. 13:925387. Doi: 10.3389/fphar.2022.925387

Copena D, Pérez-Neira D, Vázquez AM, Simón X (2022). Community forest and mushrooms: Collective action initiatives in rural areas of Galicia. Forest Policy and Economics, 135, 102660. <https://doi.org/10.1016/j.forpol.2021.102660>.

Cotter T (2014). Organic Mushroom Farming and Mycoremediation: Simple to Advanced and Experimental Techniques for Indoor and Outdoor Cultivation; Chelsea Green Publishing: White River Junction, VT, USA.

Dissanayake AA, Mills GL, Bonito G, Rennick B, Nair MG (2021). Chemical composition and anti-inflammatory and antioxidant activities of extracts from cultivated morel mushrooms, species of genus *Morchella* (Ascomycota). Int. J. Med. Mushrooms 23, 73–83. Doi: 10.1615/IntJMedMushrooms.2021039297

Dorr E, Maximilien Koegler, Benoît Gabrielle, Christine Aubry C (2021). Life cycle assessment of a circular, urban mushroom farm. Journal of Cleaner Production, 288, 125668. <https://doi.org/10.1016/j.jclepro.2020.125668>.

El-Ramady H, Llanaj X, Prokisch J (2021). Edible Mushroom Cultivated in Polluted Soils and its Potential Risks on Human Health: A short communication. Egypt. J. Soil. Sci. 61, (3), 381 – 389. DOI: 10.21608/ejss.2021.106452.1478

El-Ramady, H.; Abdalla, N.; Badgar, K.; Llanaj, X.; Törös, G.; Hajdú, P.; Eid, Y.; Prokisch, J (2022b). Edible Mushrooms for Sustainable and Healthy Human Food:

- Nutritional and Medicinal Attributes. Sustainability, 14, 4941. <https://doi.org/10.3390/su14094941>
- El-Ramady, H.; Abdalla, N.; Fawzy, Z.; Badgar, K.; Llanaj, X.; Törös, G.; Hajdú, P.; Eid, Y.; Prokisch, J (2022a). Green Biotechnology of Oyster Mushroom (*Pleurotus ostreatus* L.): A Sustainable Strategy for Myco-Remediation and Bio-Fermentation. Sustainability, 14, 3667. <https://doi.org/10.3390/su14063667>
- El-Ramady, H.; Törös, G.; Badgar, K.; Llanaj, X.; Hajdú, P.; El-Mahrouk, M.E.; Abdalla, N.; Prokisch, J (2022c). A Comparative Photographic Review on Higher Plants and Macro-Fungi: A Soil Restoration for Sustainable Production of Food and Energy. Sustainability, 14, 7104. <https://doi.org/10.3390/su14127104>
- Elsakhawy, T.; Omara, A.E.-D.; Abowaly, M.; El-Ramady, H.; Badgar, K.; Llanaj, X.; Törös, G.; Hajdú, P.; Prokisch, J (2022). Green Synthesis of Nanoparticles by Mushrooms: A Crucial Dimension for Sustainable Soil Management. Sustainability, 14, 4328. <https://doi.org/10.3390/su14074328>
- Fawzy ZF, El-Ramady H, Abd El-Fattah DA, Prokisch J (2022). Sustainable Applications of Mushrooms in Soil Science: A Call for Pictorial and Drawn Articles. Egypt. J. Soil Sci. 62 (2), 155 – 167. DOI: 10.21608/EJSS.2022.148638.1514.
- Hajdú P, Fawzy ZF, El-Ramady H, Prokisch J (2022). Edible Mushroom of *Lentinula* spp.: A Case Study of Shiitake (*Lentinula edodes* L.) Cultivation. Environ. Biodiv. Soil Security, 6, 41 – 49. DOI: 10.21608/jenvbs.2022.121848.1164.
- Jayachandran M, Xiao J, Xu B (2017). A Critical Review on Health Promoting Benefits of Edible Mushrooms through Gut Microbiota. Int J Mol Sci. 18(9), 1934. Doi: 10.3390/ijms18091934.
- Leong YK, Varjani S, Lee DJ, Chang JS (2022). Valorization of spent mushroom substrate for low-carbon biofuel production: Recent advances and developments. Bioresource Technology, 363, 128012. <https://doi.org/10.1016/j.biortech.2022.128012>.
- Meena B, Sivakumar V, Praneetha S (2020). Prospects of biodiversity and distribution of mushroom fungi in India. GSC Biol. Pharm. Sci. 13, 078–085.
- Moumita S, Das B (2022). Assessment of the prebiotic potential and bioactive components of common edible mushrooms in India and formulation of synbiotic microcapsules. LWT, 156, 113050. <https://doi.org/10.1016/j.lwt.2021.113050>.
- Okuda Y (2022). Sustainability perspectives for future continuity of mushroom production: The bright and dark sides. Front. Sustain. Food Syst. 6:1026508. Doi: 10.3389/fsufs.2022.1026508
- Pereira de Oliveira A, Naozuka J, Figueroa JAL (2023). Feasibility study for mercury remediation by selenium competition in *Pleurotus mushrooms*. Journal of Hazardous Materials, 451, 131098. <https://doi.org/10.1016/j.jhazmat.2023.131098>.
- Rahman H, Faruq MO, Abdul Hai TB, Rahman W, Hossain MM, Hasan M, Islam S, Moinuddin M, Islam MT, Azad MM (2022). IoT enabled mushroom farm automation with Machine Learning to classify toxic mushrooms in Bangladesh. Journal of Agriculture and Food Research, 7, 100267. <https://doi.org/10.1016/j.jafr.2021.100267>.
- Raman J, K-Y Jang, Y-L Oh, M Oh, J-H Im, H Lakshmanan, V Sabaratnam (2021). Cultivation and Nutritional Value of Prominent *Pleurotus* spp.: An Overview. Mycobiology, 49:1, 1-14, DOI: 10.1080/12298093.2020.1835142
- Sahithya K, Mouli T, Ankita Biswas, Mercy Scorlet T (2022). Remediation potential of mushrooms and their spent substrate against environmental contaminants: An overview. Biocatalysis and Agricultural Biotechnology, 42, 102323. <https://doi.org/10.1016/j.bcab.2022.102323>.
- Stilinovic N, Capo I, Vukmirovic S, Raškovic A, Tomas A, Popovic M, et al. (2020). Chemical composition, nutritional profile and *in vivo* antioxidant properties of the cultivated mushroom *Coprinus comatus*. R. Soc. Open Sci. 7:200900. Doi: 10.1098/rsos.200900
- Su CH, Lu MK, Lu TJ, Lai MN, Ng LT (2020). A (1→6)-Branched (1→4)-β-D-Glucan from *Grifola frondosa* Inhibits Lipopolysaccharide-Induced Cytokine Production in RAW264. 7 Macrophages by Binding to TLR2 Rather than Dectin-1 or CR3 Receptors. J. Nat. Prod. 83, 231–242. Doi: 10.1021/acs.jnatprod.9b00584
- Törös G, El-Ramady H, Prokisch, József (2022). Edible Mushroom of *Pleurotus* spp.: A Case Study of Oyster Mushroom (*Pleurotus ostreatus* L.). Env. Biodiv. Soil Security, 6, 51–59. DOI: 10.21608/jenvbs.2022.117554.1161
- Usman M, Murtaza G, Ditta A (2021). Nutritional, medicinal and cosmetic value of bioactive compounds in button mushroom (*Agaricus bisporus*): a review. Appl. Sci. 11:5943. Doi: 10.3390/app11135943
- Wang F, Fang Y, Wang L, Xiang H, Chen G, Chang X, Liu D, He X, Zhong R (2022). Effects of residual monensin in livestock manure on nitrogen transformation and microbial community during “crop straw feeding-substrate fermentation-mushroom cultivation” recycling system. Waste Management, 149, 333-344. <https://doi.org/10.1016/j.wasman.2022.06.015>.
- Wang M, Zhao R (2023). A review on nutritional advantages of edible mushrooms and its industrialization development situation in protein meat

- analogues. *Journal of Future Foods*, 3 (1), 1-7. <https://doi.org/10.1016/j.jfutfo.2022.09.001>.
- Yan P (2023). Boreal Forest Mushrooms. <https://www.ecofriendlyincome.com/boreal-forest-mushrooms> Accessed on 19.04.2023
- Yang ZY, Wang XJ, Cao Y, Dong QE, Tong JY, Mo MH (2023). Vermicomposting of *Pleurotus eryngii* spent mushroom substrates and the possible mechanisms of vermicompost suppressing nematode disease caused by *Meloidogyne incognita*. *Heliyon*, 9, 4, e15111. <https://doi.org/10.1016/j.heliyon.2023.e15111>.
- Ye D, Hu Q, Bai X, Zhang W, Guo D (2023). Increasing the value of *Phragmites australis* straw in a sustainable integrated agriculture model (SIAM) comprising edible mushroom cultivation and spent mushroom substrate compost. *Science of The Total Environment*, 869, 161807. <https://doi.org/10.1016/j.scitotenv.2023.161807>.
- Zhang Z, Zhao L, Qu H, Zhou H, Yang H, Chen H (2022). Physicochemical characterization, adsorption function and prebiotic effect of chitin-glucan complex from mushroom *Coprinus comatus*. *International Journal of Biological Macromolecules*, 206, 255-263. <https://doi.org/10.1016/j.ijbiomac.2022.02.152>.
- Zou G, Nielsen JB, Wei Y (2023). Harnessing synthetic biology for mushroom farming. *Trends in Biotechnology*, 41, Issue 4, 480-483. <https://doi.org/10.1016/j.tibtech.2022.10.001>.